

INSTRUCTIONS TO AUTHORS

1. General Information

1.1. *Biochemistry (Moscow)* is a monthly international journal published by the Russian Academy of Sciences and the Biochemical Society of Russia **concurrently** in Russian and English by IAPC Nauka/Interperiodica Publishing and Kluwer Academic/Plenum Publishers.

1.2. *Biochemistry (Moscow)* publishes papers in all fields of biochemistry and related areas (including molecular biology, bioorganic chemistry, microbiology, physiology, and medicinal biochemistry).

1.3. **Regular papers** may include final original results obtained experimentally, descriptions of new experimental methods of value for biochemistry, and theoretical material suggesting new principles and approaches to biochemical problems.

Manuscripts dealing with particularly significant and novel findings are published as **Short communications** (publication time of 4 months). The accompanying letter must give the reasons why the authors believe that the work justifies accelerated publication.

The journal also publishes **reviews** on timely topics of biochemistry and related fields, which are solicited, or suggested by the authors and approved by the Editorial Board.

Short, vivid **mini-reviews** are published in the "**Biochemistry News**" section.

The "**Discussions**" section provides an opportunity to comment on or criticize work previously published in *Biochemistry (Moscow)*, or to present a new hypothesis (maximum 4 typewritten pages). Replies from other interested parties may be solicited to polemical papers presented in this section.

In addition, the journal publishes "**Chronicles**" of congresses, meetings, and conferences with short presentations of most sensational and timely reports.

1.4. The English version of the journal is surveyed by Chemical Abstracts, Chemical Titles, Current Contents, Science Citation Index, Excerpta Medica, Index Medicus (MEDLINE), Pubmed, Biological Abstracts (BIOSIS), International Abstracts of Biological Sciences, Agricola, and Index Internacional de Cardiologia.

1.5. Beginning with the year 1996, the journal has a constantly updated WEB site (<http://www.protein.bio.msu.ru/biokhimiya>), providing the tables of contents for all journal issues together with summaries, key words and authors' addresses. Free access is also provided to 2-3

whole papers (i.e., with full text, figures, tables, etc.) of each issue, most highly ranked by the reviewers, and special journal issues published in 1997-2003 and devoted to the most timely problems of biochemistry.

1.6. Beginning with the year 2003, the journal WEB site includes the section "*Papers in Press*", allowing pre-publication access to manuscripts which were most highly ranked by reviewers.

2. Submission of Manuscripts

2.1. Manuscripts may be submitted in printed form and should be accompanied by their electronic versions (please check the diskette for computer viruses before submission) or transmitted via E-mail.

2.2. All printed materials, including manuscript body, figures, figure legends (grouped together on a separate sheet), tables, and references should be submitted **in duplicate**.

On a separate page provide the names of all authors and their postal addresses, telephone and fax numbers, and E-mail addresses and indicate the corresponding author.

Number all sheets of the manuscript in succession, including tables, references, and figure legends and indicate in the text where figures/tables should be placed.

2.3. The electronic version may be submitted on 3.5'' diskettes or CD disks as one text file with entire text, including summary, references, figure legends, tables, etc., and separate figure files.

2.4. Manuscripts may be submitted via E-mail as attachments to one of the following addresses: **bioch@maik.ru** and **ozrina@bio.chem.msu.ru**. The files should be prepared as described above. Large files should be archived with ZIP.

2.5. Regardless the way of submission, the manuscripts should be accompanied by a covering letter, stating that the submitted material has not been published or submitted elsewhere.

3. Preparation of Manuscripts

3.1. Submitted manuscripts should be condensed to the utmost compatible with clarity, but should contain sufficient detail to understand and reproduce the work.

3.2. The manuscript is to be arranged in the following order: 1) title; 2) authors' initials and last names; 3) complete names of institutions with their addresses, fax numbers and E-mail addresses; 4) abstract; 5) key words; 6) paper body.

The *Title* should be as short and informative as possible and should not contain abbreviations.

If the work was carried out at more than one institution, the authors' names should be followed by superscript numbers indicating authors' affiliations. Indicate by asterisk following the superscript number the author to whom to address correspondence.

Complete *names of institutions* with their postal addresses, fax numbers, and E-mail addresses should be given for each author.

The *Abstract* (no more than 250 words) should succinctly and clearly present the major significant results of the investigation and conclusions therefrom.

The list of *key words* should contain no more than 7 items.

A short *Running Title* should be given in a separate line after the key words.

Paper body should be divided into: 1) introductory part; 2) *Materials and Methods*; 3) *Results*; 4) *Discussion* (if discussion is short, the *Results* and *Discussion* sections may be combined); 5) *References*.

The introductory part should state the purpose of the investigation with imperative references to relevant previous works.

The description of the methods in *Materials and Methods* should be as short as possible but adequate for repetition of the work by a qualified investigator. This section should also contain the names of the manufacturers (including country name) of main materials and reagents. Only new procedures should be described in detail; published procedures should merely be referred to by literature citation. If the procedure is not generally known, it is appropriate to mention the principle and the author(s). References like "nuclease was assayed by method [7]" or "...according to [7]" should be avoided.

The *Results* are usually presented in figures and tables; some results that do not need documentation can be given in the text. Extensive discussion should not be given in this section and should be limited to an explanation of the logical links between the described experiments.

The *Discussion* should deal with interpretation and not with recapitulation of the results. Use of a simple and pictorial scheme to illustrate the major results is encouraged.

Acknowledgments of financial support should be given at the end of the *Discussion* section.

The list of *References* (see below for the style) should be as brief as possible but should contain all relevant recent publications of fundamental importance. The references should be cited in the text by numbers in square

brackets consecutively in the order of appearance in the manuscript.

3.3. Manuscript format:

3.3.1. **Regular papers** shall not exceed 20 typewritten pages, including references, tables, and figures (three figures are equivalent to one page) and 8 figures and tables. The maximum size of **short communications** is 12 typewritten pages and 4 figures or tables; of **mini-reviews**, 16 pages and 5 figures; of **reviews**, 35 pages, including references, etc.; and that of contributions to the "**Discussions**" section, 4 pages.

3.3.2. Manuscripts should be typewritten and 1.5-spaced throughout in one column without right justification on one side of 210 × 297 mm (A4 format) white paper, with a 4-cm margin on the left side. Each page should contain no more than 30 lines.

In the electronic version, text files should be submitted in Microsoft Word 6.0 or a later version, using Times New Roman font of 12 pitch size.

The text should be in one column without division into syllables. Use of italics, bold, sub- and superscripts, Greek, and mathematical symbols should adhere to journal style.

Text format should be as simple as possible, **without preprogrammed titles, insertions and cross-references, and without increased space between lines or characters. Use only Word template "Normal"**. In particular, this refers to the *References* section, as preprogrammed reference numbers are not recognized by the publisher's software.

3.3.3. Each **table** should have a title and be typed on a separate sheet. The same data will not be published in two forms, e.g., a table and a figure or a table and in the text. All table columns should have brief headings. Avoid columns with data that are easily derived from other columns, e.g., by subtraction or taking percent.

3.3.4. **Figures** may be prepared with any editor, but the final file should be in one of the following formats: *.WMF, *.TIF, *.BMP, *.DOC, *.JPG, or *.XLS.

Graph axes and curves should be appropriately labeled. If the graph contains more than one curve, they should be numbered, and explanations should be provided for each number in the figure legend. The preferred symbols for experimental points are filled and empty circles, squares, triangles, and diamonds. Individual curves may be distinguished by using solid and dashed lines. All lines, curves, and symbols should be drawn clearly and of size that allows for a required reduction or magnification on final printing. The figures should be preferably somewhat larger than the actual size in the journal but not larger than A4 format.

All figures should be identified on the back in pencil with the first author's name and the figure number.

Each figure should be supplemented with an informative legend that make its meaning comprehensible without reference to the text. Conditions specific to a partic-

ular experiment should be stated. Reference to the text is permissible to avoid repetitions and ambiguity.

Half-tone photographs and drawings should be on white glossy paper.

Simple bar histograms will not be published. The information can be given more concisely as a table or several sentences in the text.

Presentation of amino acid, nucleotide, and other sequences often requires exact vertical positioning of the elements. To reduce the chance of error and avoid tedious proofreading, the authors should submit such material in a camera-ready form.

Long mathematical expressions should be submitted as figures.

3.3.5. References containing the last names and initials of all authors should be typed on separate sheets. The style used for citation of journals, monographs, multi-author books, and theses is given in the following examples:

1. Gladysheva, I. P., Zamolodchikova, T. S., Sokolova, E. A., and Larionova, N. I. (1999) *Biochemistry (Moscow)*, **64**, 1244-1249.
2. Rodrigues Macedo, M. L., Machado Freire, M. G., Cabrini, E. C., Toyama, M. H., Novello, J. C., and Marangoni, S. (2003) *Biochim. Biophys. Acta*, **1621**, 170-182.
3. Klesov, A. A., and Berezin, I. V. (1980) *Enzyme Catalysis* [in Russian], MGU Publishers, Moscow, pp. 111-164.
4. Ryan, C.A. (1981) in *The Biochemistry of Plants* (Marcus, A., ed.) Vol. 6, Academic Press, New York, pp. 351-370.
5. Gendrolis, A. A., Serebryannikov, N. V., and Gandel', V. G. (1978) in *Prostaglandins* (Azhgikhin, I. S., ed.) [in Russian], Meditsina, Moscow, pp. 332-347.
6. Walsh, M. P. (1985) in *Calcium and Cell Physiology* (Martel, D., ed.) Springer-Verlag, Berlin, pp. 170-203.
7. Gandelman, O. A. (1992) *Kinetics and Mechanism of Bioluminescent Oxidation of Fire-Fly Luciferin*: Author's abstract of Candidate's (doctoral) dissertation [in Russian], Moscow State University, Moscow.

3.3.6. The International System of Physical Units (SI) is preferred.

3.3.7. Chemical, physical, mathematical symbols in the text, organic compound structures, and mathematical equations should be computer-printed. Letters of similar appearance in lower and upper cases (i.e., *P* and *p*, *C* and *c*, *K* and *k*), as well as those printed in italics should be clearly identified.

A product or quotient of two units should be written as in the example mol/sec (mol per second). In more complex groupings, the solidus should be combined with parenthesis to avoid ambiguity: *a/(bc)* but not *a/b/c* or *a/bc*; (*a/b*)*c* but not *a/b·c*. Use of powers is also recommended: mol·sec⁻¹. Expressions of the type mA/gel and μmol/min·mg protein are not permitted and should be substituted by mA per gel and μmol/min per mg protein.

3.3.8. The recommendation of the Nomenclature Committee of the International Union of Biochemistry should be followed for abbreviations and symbols. The use of the abbreviations given below is mandatory, and they need not be defined in the text. Other abbreviations should be defined together in on the title page under the heading "Abbreviation(s)".

Abbreviations are hindrances to readers, and their use should be restricted to the minimum. Clarity and lack of ambiguity are more important than brevity. On the other hand, it is sometimes convenient to use abbreviations for the names of substances, particularly in equations, tables, and figures.

Names of complex chemical compounds should be carefully verified. It may be better to use short formulae instead of longer names, e.g., NaCl in place of sodium chloride, CH₃COOH or AcOH in place of acetic acid. When abbreviations for chemical compounds are needed, maximum use should be made of standard chemical symbols (C, H, O, P, S, Na, Cl, etc.), trivial names (e.g., folate), and symbols (e.g., Me, Pr, Ac for methyl, propyl, acetyl, respectively).

One-letter symbols are preferred over three-letter symbols for amino acid residues in polypeptides and proteins:

Alanine	Ala	A
Arginine	Arg	R
Asparagine	Asn	N
Aspartic acid	Asp	D
Aspartic acid or asparagine	Asx	B
Cysteine	Cys	C
Glutamine	Gln	Q
Glutamic acid	Glu	E
Glutamic acid or glutamine	Glx	Z
Glycine	Gly	G
Histidine	His	H
Isoleucine	Ile	I
Leucine	Leu	L
Lysine	Lys	K
Methionine	Met	M
Phenylalanine	Phe	F
Proline	Pro	P
Serine	Ser	S
Threonine	Thr	T
Tryptophan	Trp	W
Tyrosine	Tyr	Y
Valine	Val	V

Macromolecular compounds of repeated sequences may be represented by the prefix "poly" or the subscript *n*. Thus poly(Lys) or (Lys)_{*n*} is polylysine, poly(Ala-Lys) or (Ala-Lys)_{*n*} is a polymer consisting of alanine and lysine in regular alternating sequence, and poly(Ala,Lys) or (Ala,Lys)_{*n*} is the irregular random polymer of these amino

acids. The subscript n may be replaced by an average number (e.g., (Lys)₁₀) or a range (e.g., (Lys)₈₋₁₂).

Particular amino acid residues in proteins should be designated as Arg25, Pro49, Leu54, etc.

Three-letter designations of genes should follow international rules: italicized lower-case for bacterial and mutant yeast genes and italicized upper-case for dominant yeast genes.

The symbols for sugars are as follows:

Arabinose	Ara	Glucose	Glc
2-Deoxyribose	dRib	Mannose	Man
Fructose	Fru	Ribose	Rib
Fucose	Fuc	Xylose	Xyl
Galactose	Gal		

When it is necessary to indicate furanose or pyranose, the saccharide symbol should be followed by the letter *f* or *p*: e.g., Rib*f* for ribofuranose.

The symbols for nucleosides, nucleotides, and polynucleotides are as follows:

Adenosine	A
Guanosine	G
Inosine	I
Ribosylthymine	T
Uridine	U
Xanthosine	X
Adenosine-5'-mono-, di-, and triphosphates	AMP, ADP, ATP
Cytidine-5'-mono-, di-, and triphosphates	CMP, CDP, CTP
Guanosine-5'-mono-, di-, and triphosphates	GMP, GDP, GTP
Orotidine-5'-mono-, di-, and triphosphates	OMP, ODP, OTP
Ribothymidine-5'-mono-, di-, and triphosphates	rTMP, rTDP, rTTP
Uridine-5'-mono-, di-, and triphosphates	UMP, UDP, UTP

The corresponding deoxyribonucleotides are designated by the same symbol preceded by the low-case letter "d", e.g., dATP, dGTP, etc.

AMP isomers are designated as 2'-AMP, 3'-AMP, 5'-AMP, 3':5'-AMP (adenosine-3':5'-monophosphate, cAMP).

The following symbols are used for specific preparations of nucleic acids:

Deoxyribonucleic acid	DNA
Complementary DNA	cDNA
Mitochondrial DNA	mtDNA
Ribonucleic acid	RNA
Mitochondrial RNA	mtRNA

Messenger RNA	mRNA
Ribosomal RNA	rRNA
Transfer RNA	tRNA
Specific tRNA	tRNA ^{Ala} , tRNA ^{Glu} , etc.
Isoacceptor tRNA	tRNA ₁ , tRNA ₂ , etc.
Aminoacylated tRNA	Ala-tRNA, Glu-tRNA, etc.

Polyphosphoinositides and their hydrolysis products should be designated in the following way:

Phosphatidyl	Ptd
Inositol	Ins
Phosphate	P

Thus, PtdIns(4,5)P₂ stands for phosphatidylinositol 4,5-bisphosphate.

Names of enzymes may be abbreviated, e.g., G6PDG (glucose-6-phosphate dehydrogenase). The abbreviation should be defined in the "Abbreviations" paragraph on the title page. Substrate name used as part of the trivial enzyme name may be abbreviated, e.g., ATPase, Glu-decarboxylase.

The following abbreviations may be used without definition:

acyl-CoA	acyl derivative of coenzyme A
BSA	bovine serum albumin
CM-cellulose	carboxymethyl cellulose
CoA, CoASH	coenzyme A
DEAE-cellulose	diethylaminoethyl cellulose
SDS	sodium dodecyl sulfate
TCA	trichloroacetic acid
EDTA	ethylenediaminetetraacetate
EGTA	ethylene glycol-bis(β-aminoethyl ether) N,N,N', N'-tetraacetate
FAD, FADH ₂	flavin-adenine dinucleotide and its reduced form
FMN, FMNH ₂	riboflavin-5'-phosphate and its reduced form
GSH, GSSG	reduced and oxidized glutathione
G-protein	guanine-nucleotide-binding regulatory protein
IgG	immunoglobulin G
NAD, NAD ⁺ , NADH	nicotinamide-adenine dinucleotide and its oxidized and reduced forms
NADP, NADP ⁺ , NADPH	nicotinamide-adenine dinucleotide phosphate and its oxidized and reduced forms
PAGE	polyacrylamide gel electrophoresis
P _i	orthophosphate
PP _i	pyrophosphate
POPOP	1,4-bis(5-phenyl-2-oxazolyl) benzene

PPO	2,5-diphenyloxazol
Q, QH ₂	ubiquinone, ubiquinol

Class names (fatty acids, protein, virus, etc.) and short names (folate, furan, etc.) **are not abbreviated**. **Abbreviations should not be used** for terms such as “central nervous system”, “red blood cells”, or “extracellular fluid” and for names of tissue preparations, buffers, and suspension media.

The following abbreviations may be used for common physicochemical methods and related terms: CD, circular dichroism; EPR, electron paramagnetic resonance; ESR, electron spin resonance; GLC, gas-liquid chromatography; HPLC, high-pressure liquid chromatography; IR- and UV-spectroscopy, infrared and ultraviolet spectroscopy; NMR, nuclear magnetic resonance; ORD, optical rotary dispersion; SDS-PAGE, polyacrylamide gel electrophoresis in the presence of sodium dodecyl sulfate; TLC, thin-layer chromatography.

3.3.9. Nomenclature of isotopically labeled compounds. The isotope symbol is placed in square brackets directly attached to the front of the name: [¹⁴C]urea, [α -¹⁴C]leucine, L-[methyl-¹⁴C]methionine. When more than one position in a substance is labeled and the positions are not indicated, the number of labeled atoms is shown by the subscript on the right-hand side of the symbol: [¹⁴C₂]glycolic acid. The symbol “U” indicates uniform labeling (in [U-¹⁴C]glucose each molecule has ¹⁴C at all six positions), and symbol “G” indicates general labeling (in [G-¹⁴C]glucose ¹⁴C may be present at any, but not necessarily all, of the six positions). In the latter case, [¹⁴C]glucose will suffice.

The isotope prefix precedes the part of the compound name to which it refers: iodo[¹⁴C]acetic acid, 1-amino-[¹⁴C]methylcyclopentanol (H₂N¹⁴CH₂C₅H₈OH), fructose 1,6-[1-³²P]bisphosphate. Terms such as “¹³¹I-labeled albumin” should not be contracted to “[¹³¹I]albumin” since native albumin does not contain iodine; however, “[¹³¹I]iodoalbumin” is acceptable.

When compound contains isotopes of more than one element, their symbols are arranged alphabetically [3-¹⁴C, 2,3-D¹⁵N]serine. Deuterium may be designated by ²H or D, tritium by ³H or T.

The positions of isotope labeling are indicated by Arabic numerals, Greek letters, or prefixes placed within the square brackets before the element symbol to which they are attached by a hyphen: [1-³H]ethanol, L-[α -¹⁴C]leucine, [carboxyl-¹⁴C]leucine, [3,4-¹⁴C, ³⁵S]methionine, L-[methyl-¹⁴C]methionine.

The above rules also apply when the labeled compounds are designated by standard abbreviations or symbols: [α -³²P]ATP, [³²P]CMP (not CM³²P!). Labeled orthophosphate and pyrophosphate may be designated by ³²P_i and ³²PP_i, respectively.

Square brackets may be omitted for simple molecules by writing their chemical formulae: ¹⁴CO₂, H₂¹⁸O, D₂O,

H₂³⁵SO₄, ³²PO₄³⁻ (but [³²P]phosphate). The square brackets are not to be used when the isotopic symbol is attached to a word that is not a chemical name or refers to a class name of compounds: ¹³¹I-labeled, ³H-ligand, ¹⁴C-steroids, ¹⁴C-amino acids.

When describing experiments with labeled compounds, absolute values of the radioactivity should be given, wherever possible, in curies (Ci), becquerels (Bq), disintegrations per minute and second (dpm and cpm, respectively), or pulses per minute (pulse/min).

3.3.10. Recommendations on specific topics common for biochemical literature are given below (see also *Biochem. J.*, **289**, 1-15 (1993)).

Animals, plants, microorganisms. The full binomial names should be included for all experimental animals (other than common laboratory animals) and plants. The strain, the variety, and, if possible, the source of the material should be given. Reports describing effects of changes in feeding should contain the compositions of the feeding material (growing media).

In the title, the summary, and at first mention in the text, **full** binomial Latin names, italicized, should be given for **microorganisms**. Elsewhere in the text, single-letter abbreviation may be given for the generic name along with full species name. The number of the organism in the collection from which the organism was obtained should be stated. Alternatively, a strain number (not italicized) should be given. If two genera with the same initial letters are studied, abbreviations such as *Strep.* and *Staph.* may be used. Ranks higher than genus (e.g., Eubacteriales, Lactobacillae) and generic names used adjectively (e.g., staphylococcal) are not italicized.

Centrifugation. When conditions for centrifugation are critical, sufficient information should be provided for the experiment to be repeated: the centrifuge rotor, the quantitative composition of the suspension medium, operation temperature, the time of rotor operation at constant velocity (ignoring acceleration and deceleration periods), the centrifugal field based on the average radius of rotation of the liquid. For example: “The centrifugation was performed for 15 min at 2°C and 10,000g (r_{av} 8 cm)”.

For density-gradient centrifugation, centrifuge and rotor manufacturer(s), temperature, and gradient composition should be stated. Results should preferably be presented as a function of distance from rotor center rather than fraction number; it is then unnecessary to indicate top and bottom of the gradient. If fraction numbers are used, the top and bottom of the gradient should be indicated.

For ultracentrifugation, the following parameters are used: sedimentation coefficient (not constant), s ; sedimentation coefficient at zero concentration at 20°C in water, $s_{20,w}^0$; Svedberg unit (10^{-13} s), S; partial specific volume, \bar{v} ; diffusion coefficient, D ; diffusion coefficient at zero concentration at 20°C in water, $D_{20,w}^0$. The tempera-

ture at which the sedimentation and diffusion measurements were made should be stated.

Chromatography. Use of photographs or schemes of paper and thin-layer chromatography should be restricted to cases when it is difficult to give corresponding information in the text, e.g., when homology is demonstrated.

The rate of movement of a substance relative to the solvent front in paper or thin-layer chromatography is characterized by R_f value. Solvent composition is best described in the form butan-1-ol—CH₃COOH—H₂O (4 : 4 : 1 v/v).

Elution diagrams for column chromatography should be shown with the effluent volume increasing from left to right. Units of concentration and volume should be shown clearly. Column dimensions and, if possible, column void volume (V_0) should also be stated. Elution peak maximum may be characterized by elution volume (V_e) or, preferably, by partition coefficient (α or K_D). Calibration curves (e.g., plots of molecular mass versus V_e or K_D) for columns will not be published.

Electrophoresis. Photographs of gel electrophoregrams will be published provided that they bear some important information; drawings or densitograms may be more informative in certain cases. The composition of the electrophoretic medium, pH, temperature, electrophoretic mobilities (m), and operative voltage should be quoted. The symbol pI should be used for isoelectric point.

Enzymes. For nomenclature, the recommendations of the latest edition of Enzyme Nomenclature (1992, Academic Press, San Diego-New York) should be followed. Units of enzyme amount should be defined in each paper in terms of the rate of the reaction catalyzed under specified conditions. The SI unit for the rate is 1 mol of substrate transformed per 1 sec (or 1 mol of product formed per 1 sec). This gives the unit of enzyme amount called katal (symbol: kat). Units of enzyme amount may be also expressed in terms of the amounts that catalyze other rates, e.g., 1 μ mol of substrate transformed per 1 min.

Concentrations of protein solutions are often measured versus a solution of a standard protein (e.g., BSA). The standard protein used, its source, and, if possible, water content should be quoted.

The rate constants for the forward and backward reactions at the n th step of a multistep enzyme-catalyzed reaction should be represented by k_n and k_{-n} , respectively. The Michaelis constant (K_m) is defined as substrate concentration ($[S]$) which corresponds to $v = V/2$, where V (or V_{\max}) is the initial rate of product appearance or substrate disappearance when the enzyme is saturated with the substrate, and v is the initial rate at a given substrate concentration. For reactions involving two substrates (A and B), $K_m^A = [A]$ when $v = V/2$ and $[B]$ is extrapolated to infinity; a value of $[A]$ at which $v = V/2$ at a finite concentration (which should be specified) of B should be

referred to as an apparent Michaelis constant for A ($K_{m,app}^A$). Other parameters used in enzyme kinetics include: K_s , dissociation constant for enzyme—substrate complex; K_i , dissociation constant for enzyme—inhibitor complex; $[I]_{50}$, inhibitor concentration at which rate is decreased by half; h , Hill coefficient (parameter in Hill equation used to describe sigmoidal v versus substrate or inhibitor concentration curves) (see also “Recommendations on Symbolism and Terminology in Enzyme Kinetics” published in *Arch. Biochem. Biophys.*, **224**, 732–740 (1983)).

Amount of substance, molecular mass, daltons, and molar concentration. The SI unit of the amount (n) of substance is mole (abbreviated mol), i.e., the amount of substance containing the same number of structural units (molecules, atoms, ions, electrons, etc.) as the number of carbon atoms contained in 0.012 kg of ¹²C. Avogadro's number $N_A = 6.02 \cdot 10^{23}$ per mol) gives the number of structural units in a mole of any substance. Molar mass (M) is the mass of 1 mol of the substance (m/n), and its dimension is g/mol or kg/mol. Mass (m , g), amount (n , mol), and molar mass (M , g/mol) are different terms which are linked to one another with the relationship $m = nM$. There are two preferred ways of specifying the mass of a biochemical entity. Relative molecular mass (M_r , formerly “molecular weight”) is the ratio of the mass of a molecule to 1/12 of the mass of the atom ¹²C. Hence, it is dimensionless. Molecular mass is the mass of one molecule of a substance expressed in daltons; the dalton is defined as 1/12 of the mass of the atom ¹²C or M/N_A . Thus, a protein may be said to have a relative molecular mass of 50,000 ($M_r = 50,000$) or a molecular mass of 50,000 daltons (better, 50 kD), and may be referred to as the 50,000- M_r protein or the 50-kD protein. It is not correct to express M_r in daltons. Either M_r or molecular mass (kD) should be used throughout the paper.

Solutions should be described in terms of molarity (M , mM, μ M, etc.), i.e., the number of moles of substance contained in 1 liter of the solution, not normality (N). The decimal system should be used, e.g., 0.25 M HCl. The term % must be defined as w/w, w/v, or v/v, e.g., 5% (w/v) means 5 g/100 ml. For aqueous solutions of less than 1%, w/v need not be stated since it is obvious that the concentration is given in terms of mass of solute. For solutions of salts, expressed in %, it should be made clear whether the compounds are hydrated or anhydrous.

Nucleotide sequences. Authors should remember that nucleotide sequence should be determined in both DNA chains. A clear description of the determination and complete sequence data will suffice.

Powers in tables and figures. Authors must exercise care in the use of powers to avoid numbers with too many digits in table headings and in figures. This is illustrated by the following examples: 1) a concentration 0.00015 M may be expressed as $15 \cdot 10^{-5}$ M but it is preferable to give it using a prefix, as 0.15 mM or 150 μ M; listing of 0.15

under the heading "Concentration, mM" or 150 under "Concentration, μM ", or 15 under "Concentration $\times 10^5$, M" are all appropriate (but not 15 under the heading "Concentration, $\text{M} \times 10^{-5}$!"); 2) listing of 2 under the heading " $10^3 k$ " means $k = 0.002$, and 2 under the heading " $10^{-3} k$ " means $k = 2000$; 3) complex quantities are treated similarly; a value of 200 M^{-1} for $1/[\text{S}]$ would appear as "2" under the heading " $10^{-2}/[\text{S}], \text{M}^{-1}$ " or as "0.2" under the heading " $1/[\text{S}], \text{mM}^{-1}$ ". Concentrations may conveniently be indicated by square brackets.

The following prefixes and symbols should be used for multiples and subdivisions of units:

Multiple	Prefix	Symbol
10^{12}	tera	T
10^9	giga	G
10^6	mega	M
10^3	kilo	k
10^2	hecto	h*
10	deca	da*
10^{-1}	deci	d*
10^{-2}	centi	c*
10^{-3}	milli	m
10^{-6}	micro	μ
10^{-9}	nano	n
10^{-12}	pico	p
10^{-15}	femto	f
10^{-18}	atto	a

* To be avoided whenever possible (except for cm).

A combination of a prefix and a unit is treated as one symbol and may be raised to a power without using brackets, e.g., mM^{-1} and cm^2 .

Buffer solutions. These must be specified in a way allowing readers to reproduce the experimental conditions. It is useful to give complete composition of each buffer solution in the *Materials and Methods* section or at the first mention, e.g., 0.09 M CH_3COONa /0.01 M CH_3COOH , pH 5.6 (which means that the buffer solution has these concentrations of these substances). A short designation "0.1 M sodium acetate buffer, pH 5.6" may be used thereafter throughout the paper. If a buffer contains two or more ionizable substances, e.g., pyridine and CH_3COOH , the concentration of each component must be specified.

Trivial names of the following common buffers may be used without definition:

Aces	2-[(Amino-2-oxoethyl)amino]ethanesulfonic acid
Ada	(N-[2-Acetamido]-2-iminodiacetic acid
Bes	2-[Bis(2-hydroxyethyl)amino]ethanesulfonic acid
Bicine	N,N-Bis-(2-hydroxyethyl)glycine

Bistris	2-[Bis-(2-hydroxyethyl)amino]-2-(hydroxymethyl)propane-1,3-diol
Hepes	4-(2-Hydroxyethyl)-1-piperazine-ethanesulfonic acid
Hepps	4-(2-Hydroxyethyl)-1-piperazine-propanesulfonic acid
Mes	4-Morpholine-ethanesulfonic acid
Mops	4-Morpholine-propanesulfonic acid
Pipes	1,4-Piperazinediethanesulfonic acid
Taps	3-{[2-Hydroxy-1,1-bis(hydroxymethyl)-ethyl]amino}-1-propanesulfonic acid
Tes	3-{[2-Hydroxy-1,1-bis(hydroxymethyl)-ethyl]amino}ethanesulfonic acid
Tricine	N-[2-Hydroxy-1,1-bis(hydroxymethyl)-ethyl]glycine
Tris	2-Amino-2-(hydroxymethyl)-1,3-propanediol

Incubation media such as Krebs–Ringer solution, Eagle's medium, or Waymouth's medium should be defined by citing the reference or by giving their compositions.

Spectra and spectroscopic data. Full spectra should be published only if they demonstrate novel or important information. The spectra for UV or visible absorption, fluorescence, circular dichroism, and optical rotation should have a wavelength scale (nm or μm). Where possible, molar terms should be used when describing absorption, optical rotation, and circular dichroism. As stated above, commonly used abbreviations of methods (ORD, CD, EPR, ESR, and NMR) need not be defined.

Visible and ultraviolet absorption spectroscopy. The value of $\log(I_0/I)$ is called attenuation, and this reduces to absorbance when scattering and reflection are negligible. If scattering is significant, e.g., when culture cell density is estimated, the more general term attenuation should be used. Otherwise, the term absorbance should be used, but not extinction or optical density. The symbols used: A , absorbance ($\log(I/I_0)$); a , specific absorption coefficient ($\text{liter} \cdot \text{g}^{-1} \cdot \text{cm}^{-1}$) (alternatively used $A_{1\text{cm}}^{1\%}$); ϵ , molar absorption coefficient (absorbance of a 1 M solution in a 1 cm light-path) ($\text{liter} \cdot \text{mol}^{-1} \cdot \text{cm}^{-1}$ or $\text{M}^{-1} \cdot \text{cm}^{-1}$ but not $\text{cm}^2 \cdot \text{mol}^{-1}$). Wavelength (nm) at which measurements are done are given without units (e.g., $A_{1\text{cm},420}^{1\%}$). No equals sign is placed between ϵ or A and its numerical value.

IR spectra are reported as percentage transmittance (T) versus wavelength (μm) or frequency (cm^{-1}).

Optical rotation is reported as the specific rotation ($[\alpha]_\lambda^t$), which is numerically equal to the rotation in degrees of a 1 g/ml solution in a 1 dm (10 cm) light-path at wavelength λ and temperature t . Solution concentration (g/100 ml) and solvent should be stated, e.g., $[\alpha]_{420}^{20}$ 27.5° (2 g/100 ml methanol). Molar expressions (molar rotation) may be also used: $[M] = [\alpha] \cdot M_r$ and $[m] = [\alpha] \cdot M_r/100$.

For biopolymers, optical rotatory dispersion ($[m]_{\text{m.r.w}}$) is reported for the mean residue (monomer) M_r ; the dimension of $[m]_{\text{m.r.w}}$ is $\text{deg} \cdot \text{cm}^2 \cdot \text{dmol}^{-1}$.

Optical rotatory dispersion is reported as the variation of $[\alpha]$ or $[m]$ with wavelength or frequency.

Circular dichroism is reported as the molar absorption coefficient ($\Delta\epsilon = \epsilon_L - \epsilon_R$, where ϵ_L and ϵ_R are absorption coefficients for the light polarized to the left and to the right) or as molar ellipticity $[\theta]_M$. For biopolymers, molar concentrations in terms of the mean residue M_r are generally used. Units of $\Delta\epsilon$ are $\text{liter}\cdot\text{mol}^{-1}\cdot\text{cm}^{-1}$ or $\text{M}^{-1}\cdot\text{cm}^{-1}$; units of $[\theta]_M$ are as for $[m]$ in terms of the mean residue. The relationship between $\Delta\epsilon$ and $[\theta]_M$ is $[\theta]_M = 3300 \Delta\epsilon$.

Fluorescence spectroscopy. In reporting fluorescence (F) excitation and emission spectra, it should be stated whether they are normalized or corrected, and what is the nature of the correction. Fluorescence-polarization data and spectra are reported as polarization ratio (P) or anisotropy ratio (A); both are dimensionless.

Statistical treatment of results. Data from a large number of independent experiments should be reported in a way permitting evaluation of their reproducibility and significance. When the goal is to determine quantitative or statistical characteristics of a population, the information is adequately given by: 1) the number of independent experiments (replicate measurements in one animal, results from pooled tissues, etc., represent only one independent estimate); 2) the mean value; 3) the standard deviation, the coefficient of variation, or the standard error of the estimate of the mean value. It should be clearly stated whether the standard deviation or the standard error is used. A convenient form for inclusion in a table is, for example, 263 ± 2.5 (10), where the number in parentheses represents the number of values used in calculating the mean.

If the results are claimed significant, a significance test should be performed and probability estimated.

Normal-distribution statistics should be used unless otherwise established.

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